

Broadmountain Winery - 8 miles from Armstrong
 Mollie Kelly - PSU
 enologist

Controlling
 Brettanomyces
 Wine yeast = *Sacharomyces*
 Prof. Charles G. Edwards
 Viticulture/Enology Team
 Washington State University
 Pullman, WA, USA

* Always rehydrate yeast.
 Add yeast to airm water.
~~water~~
 Use a microscope to
 check wine.

What is "Brett"?
 Dekkera
 Sporulating form
 Sewage
 Mousy
 Bandaid
 Honey

Sacharomyces (wine yeast) w/o
 oxygen. Non-Sac (wild yeast)
~~the~~ must have oxygen.
 Bad Yeasts:
 Klæckra
 Brettanomyces B. *bruxellensis*
 Controlled by SO₂ & filtration

What is "Brett"?
 Dekkera
 • Sporulating form
 Brettanomyces
 • Non-sporulating form
 • Yeast does not form spores
 in wine
 Different names → SAME YEAST!

comes from
 grape juice concentrate
 Zygosaccharomyces - very
 Cretis explosions. Can't be
 controlled.

Buy new met busele every year

Bacteria:

Sources

- A. Used barrels.
- B. Unfiltered wines brought into winery.
- C. Droplets in air.
 - 1. Found in winery processing areas.
(Connell et al., 2002)
- D. Found in vineyards?
 - 1. "...grape berry is the primary source..."
(Renouf and Lonvaud-Funel, 2007)

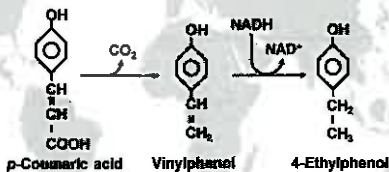
Brettanomyces is nowhere...

...yet everywhere ...

at the same time.

Linda Bisson (2018)

Synthesis of 4-Ethylphenol



Oenococcus oeni
Conducts malolactic ferm.
Desirable when expected
Undesirable when unexpected

Pediococcus

- Several species (4)
- Spoilage issues
 - Biogenic amines or ropiness
 - Bitterness
 - Excessive diacetyl (butter)
 - Dirty socks

SO₂ + filtration can control

A- Lactobacillus - 12 species

B- Many spoilage issues

① Volatile acidity

② Mousiness

③ Cotton-like fibers

④ "Stuck" fermentation

C- Grow rapidly

Acetobacter

A. Several species

B. Volatile acidity

C. Needs Oxygen for growth

What have we learned about controlling *Brettanomyces*?

- Role of SO₂?
- Ethanol x temperature?
- Removal by filtration?
- Impact of must nutrition
- Survival in oak barrels?

Management

control spoilage by using multiple "tricks"

0.8 mg/L SO₂ thought to ~~inhibit~~ inhibit microorganism.

Minimize post-alcoholic nutrients

Impact of SO₂

A. Can SO₂ cause "VBNC"?

1. "Viable but not culturable."
2. Yeast alive but will not grow on media.
3. If valid → yeast could be undetected.

B. How to detect VBNC?

1. Measure metabolic activity or qPCR(?).
2. Fluorescence microscopy.

Green = viable Orange/Red = dead

Sanitation Practices

+ Clean and sanitize equipment.
 + watch for dead spots in pipes
 + Be diligent - Monitor your wine! Use the allas-
sample wine often.

+ Run chemical & Microbiological tests

Book - Wine Microbiology

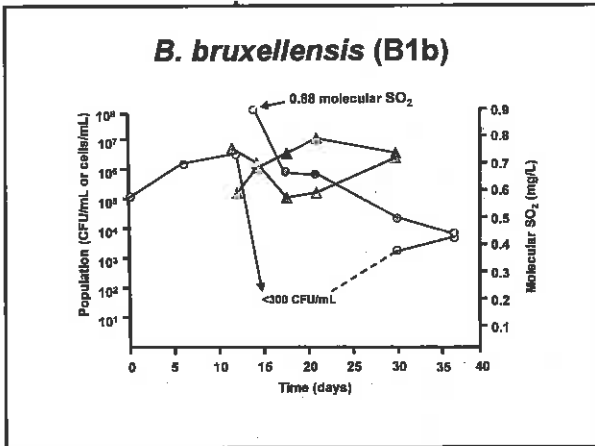
Brettanomyces

Active cell

Dead cell

Part II

SO₂



Decreasing Ph make the sulfite more effective

Is there a relationship between ethanol and temperature that could be useful to control *Brettanomyces*?

Impact of Ethanol

A. Many wine microbes can grow in 12 to 14%.

1. *Lactobacillus fructiferans* can tolerate higher amounts, approaching 16-18%.
2. Non-*Saccharomyces* yeasts have lower ethanol tolerance than wine yeast *Saccharomyces*.
 - a. 4 to 5% ethanol.
 - b. Temperature and pH greatly influence alcohol tolerance.
3. What about *Brettanomyces*?

- A. SO₂ inhibits growing
- B. Binds acetaldehydes - use SO₂ (green apple, banana)
- C. Some Microbes are resistant to SO₂ (zy.)
- D. *Saccharomyces* is susceptible to SO₂ so don't add too much

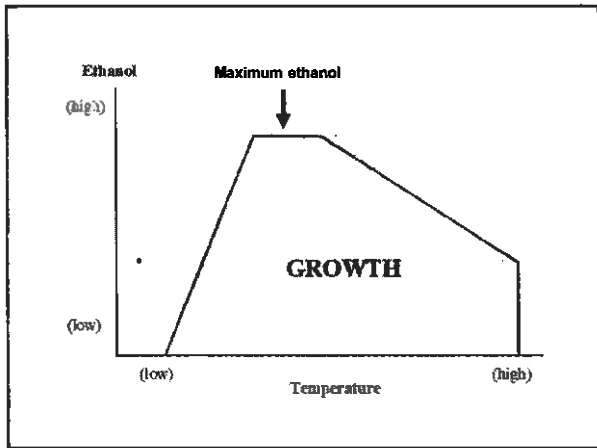
Forms of SO₂:

1. Free vs. Bound
2. Free molecular vs Sulfite vs Bisulfite

High Ph will cause spoilage.
Add "tartaric acid"

• 8 mg/l of m SO₂

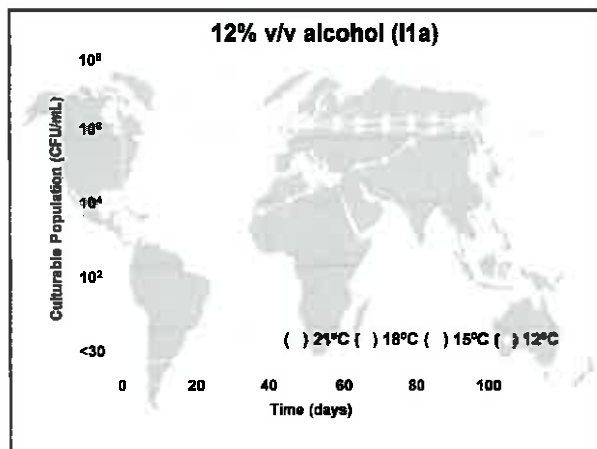
Free SO₂

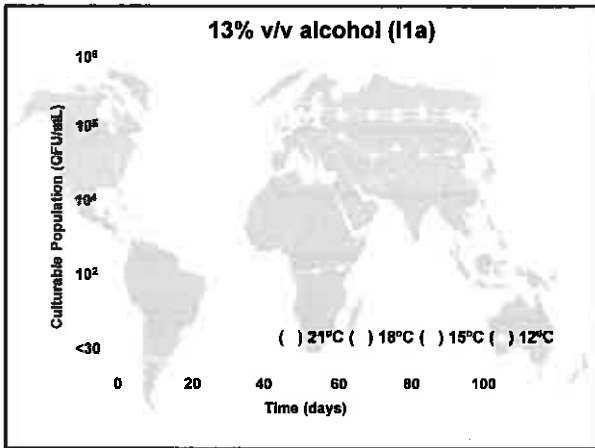


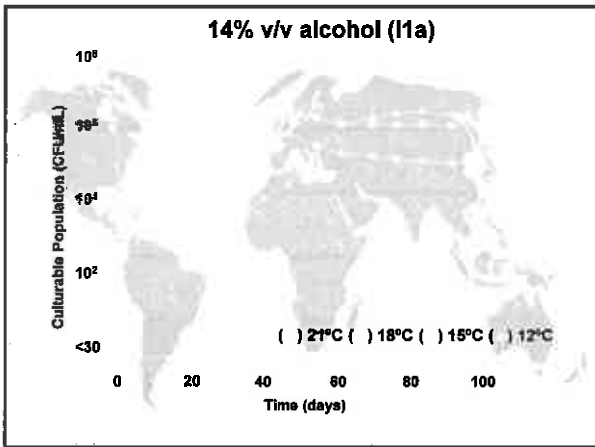
Research Focus (Ethanol x Temperature)

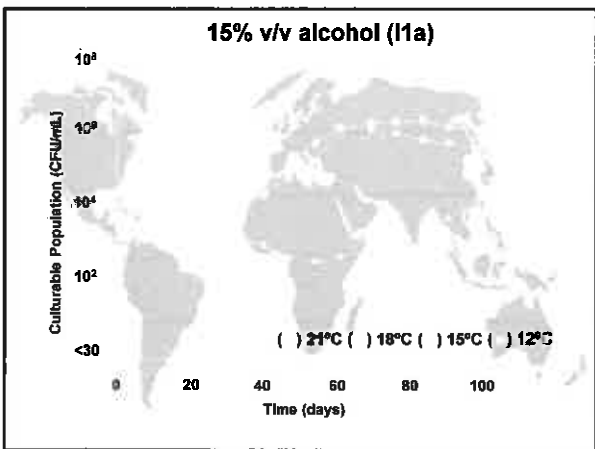
A. General protocol.

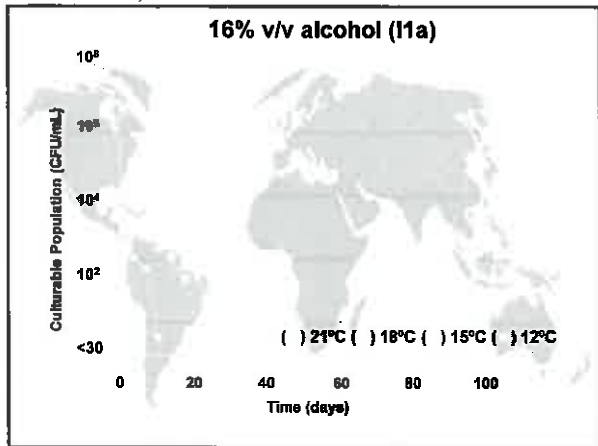
1. Inoculate Merlot wine at high populations (>10⁴ cfu/mL).
2. Experimental design.
 - a. Ethanol = 12, 13, 14, 15, or 16% v/v
 - b. Temperature = 12°, 16°, 18°, or 21° C.
 - c. Strains = F3 and I1a
3. Weekly sample wines for 100 days.
4. Measure 4-EP after 100 days.











Production of 4-Ethylphenol ($\mu\text{g/L}$)

$^{\circ}\text{C}$	Ethanol (%)	Strain F3	Strain I1a
21 $^{\circ}$	12	(+)	(+)
	13	(+)	(+)
	14	(+)	(+)
	15	(-)	(+)
	16	(-)	(-)
18 $^{\circ}$	12	(+)	(+)
	13	(+)	(+)
	14	(+)	(+)
	15	(-)	(+)
	16	(-)	(-)

(-) <math>< 30 \mu\text{g/L}</math>; (\pm) 31 to 1000 $\mu\text{g/L}</math>; (+) >1300 $\mu\text{g/L}</math>$$

Production of 4-Ethylphenol ($\mu\text{g/L}$)

$^{\circ}\text{C}$	Ethanol (%)	Strain F3	Strain I1a
18 $^{\circ}$	12	(+)	(+)
	13	(+)	(+)
	14	(+)	(+)
	15	(-)	(-)
	16	(-)	(-)
12 $^{\circ}$	12	(-)	(\pm)
	13	(-)	(-)
	14	(-)	(-)
	15	(-)	(-)
	16	(-)	(-)

(-) <math>< 30 \mu\text{g/L}</math>; (\pm) 31 to 1000 $\mu\text{g/L}</math>; (+) >1300 $\mu\text{g/L}</math>$$



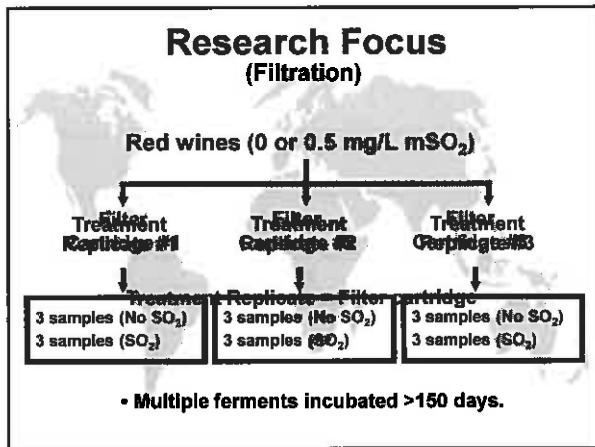
Filtration

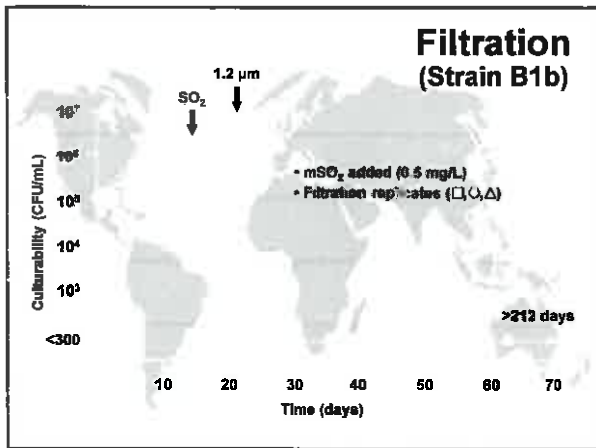
A. Desirable to minimize filtration

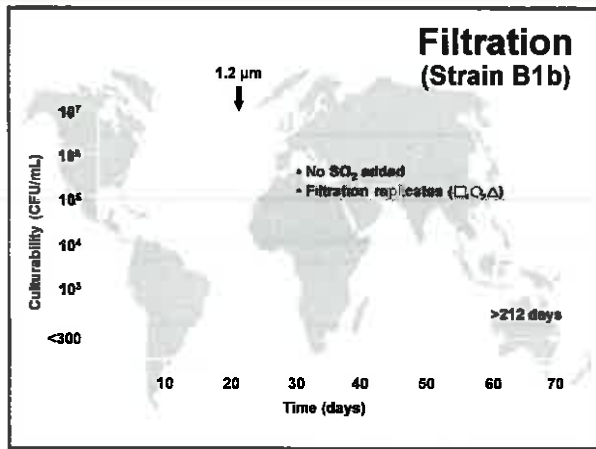
1. Expensive, quality concerns.

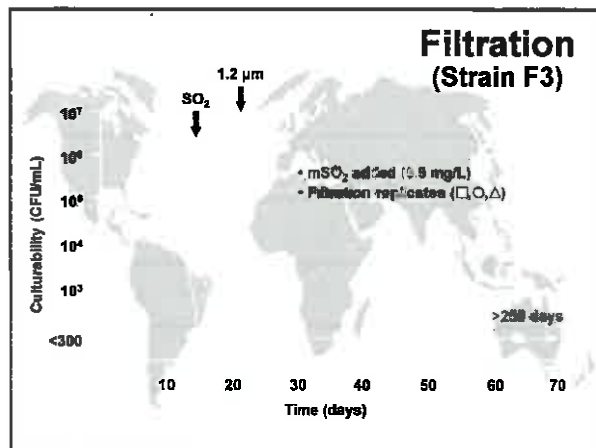
B. "VBNC" cells can exhibit dwarfing.

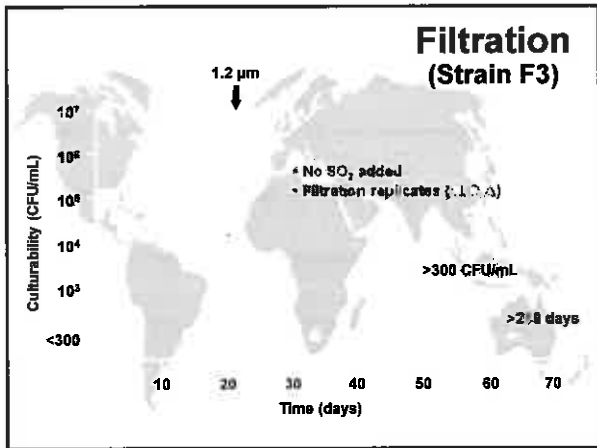
1. Cells smaller than normal.
2. Problem = filtered wines depending on pore size of membrane.

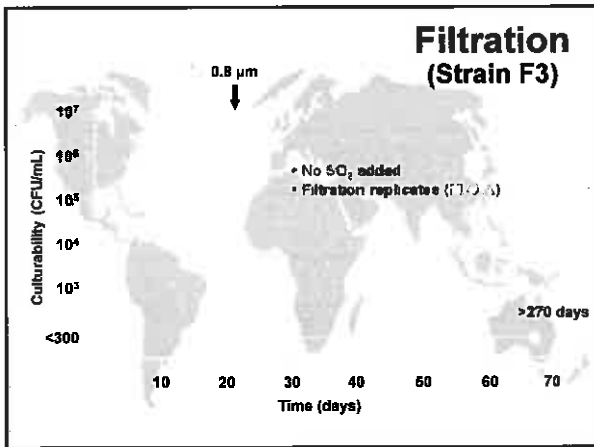












Impact of Must Nutrition

1. If more N is added to grape must → is more N present in resultant wines?
2. Does higher N in wines encourage infections?
 - What amount of N is needed to support growth?

Residual nitrogen in "synthetic" wines

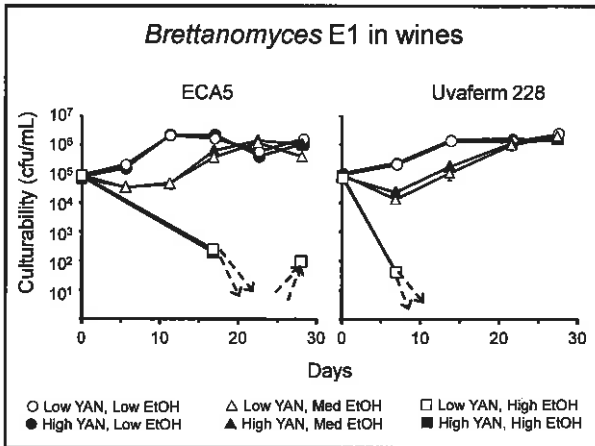
Yeast strain	Must treatment		Wine	
	YAN (mg N/L)	Sugar (g/L)	α -Amino nitrogen (mg N/L)	Ammonia nitrogen (mg N/L)
ECA5	150	230	50 ^b	1 ^b
		250	48 ^{bc}	1 ^b
		270	45 ^b	1 ^b
	250	230	85 ^a	60 ^b
		250	84 ^a	60 ^b
		270	86 ^a	60 ^b

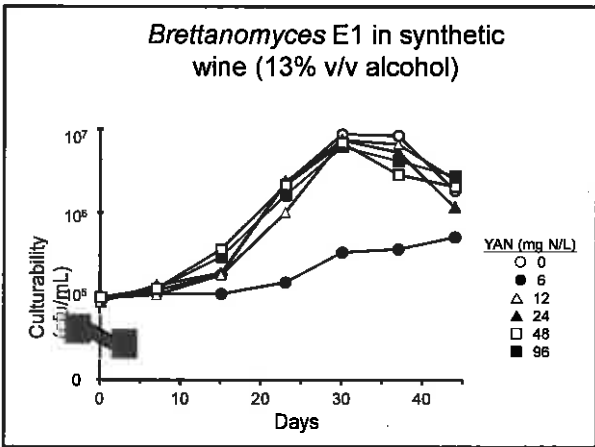
Residual nitrogen in "synthetic" wines

Yeast strain	Must treatment		Wine	
	YAN (mg N/L)	Sugar (g/L)	α -Amino nitrogen (mg N/L)	Ammonia nitrogen (mg N/L)
Uvaferm 228	150	230	45 ^b	3 ^b
		250	44 ^b	2 ^b
		270	53 ^b	3 ^b
	250	230	90 ^a	30 ^b
		250	87 ^a	30 ^b
		270	89 ^a	50 ^a

Ethanol concentrations in wines

Sugar in musts (g/L)	Ethanol in wines (% v/v)	
230	13.5 to 14.1	"low"
250	14.7 to 14.9	"medium"
270	15.6 to 15.9	"high"





Survival in Oak Barrels

Methods to eradicate *Brettanomyces* from oak?

Given chemical methods can only penetrate wood few mm, what about use of heat?

1. Important factors affecting yeast penetration.
 - a. Oak species, toasting, stave location
2. Use of steam and hot water.

Research Focus (Survival in Oak Barrels)

A. Heat destroys microbes including *Brettanomyces*.

1. Inactivation begins at 50°C.

B. Time at °C for 90% reduction in population (D-value).

1. At 55°C → 12 to 30 seconds.

• Time and temperature critical.

2. Effect of initial population.

• Higher population →
longer time and/or
higher temperature.

Experimental Design

16L Barrels (new)

American oak (light vs. heavy toast)

French oak (light vs. heavy toast)

225L Commercial Barrels (3 year old)

American oak (2 of medium-heavy toast)

French oak (2 of medium-light toast)

Experimental Design (16L Barrels)



Research Focus (Yeast Penetration)

Table saw

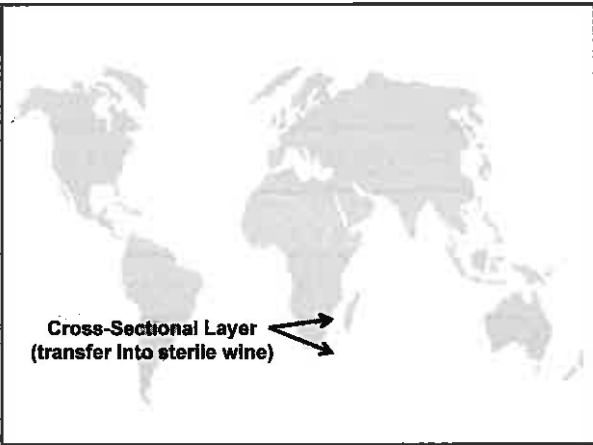
- Cut staves into 3 x 10 cm blocks

Band saw

- Saw cross-sections (4 mm thick)

Incubate cross-sections (up to 90 days)

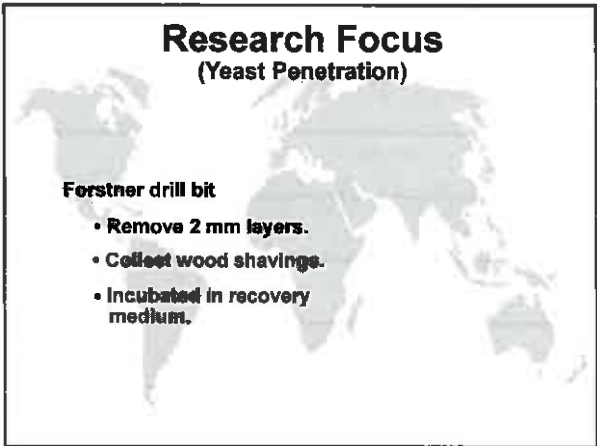
- EBB recovery medium
- Sterilized low alcohol red wine
- Advantage → large surface area



Cross-Sectional Layer
(transfer into sterile wine)

Recovery of *Brettanomyces* (225L barrels)

Research Focus
(Yeast Penetration)



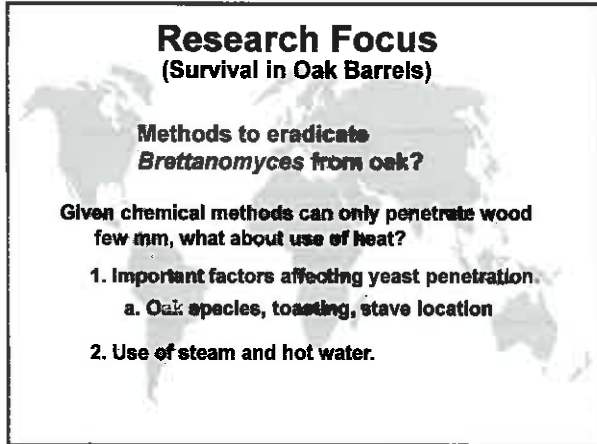
Forstner drill bit

- Remove 2 mm layers.
- Collect wood shavings.
- Incubated in recovery medium.

Research Focus
(Penetration into Oak Staves)



Research Focus
(Survival in Oak Barrels)



Methods to eradicate *Brettanomyces* from oak?

Given chemical methods can only penetrate wood few mm, what about use of heat?

1. Important factors affecting yeast penetration.
 - a. Oak species, toasting, stave location
2. Use of steam and hot water.

Research Focus (Survival in Oak Barrels)

- A. Heat destroys microbes including *Brettanomyces*.
 - 1. Inactivation begins at 50°C.
- B. Time at °C for 90% reduction in population (D-value).
 - 1. At 55°C → 12 to 80 seconds.
 - Time and temperature critical.
 - 2. Effect of initial population.
 - Higher population → longer time and/or higher temperature.

Research Focus (Survival in Oak Barrels)

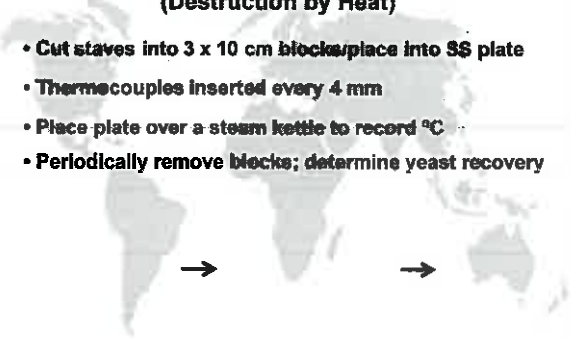


Recovery of *Brettanomyces* (16L barrels; top staves)

**Recovery of *Brettanomyces*
(16L barrels; bottom staves)**

**Research Focus
(Destruction by Heat)**

- Cut staves into 3 x 10 cm blocks/place into SS plate
- Thermocouples inserted every 4 mm
- Place plate over a steam kettle to record °C
- Periodically remove blocks; determine yeast recovery



Oak barrel staves (16L)

American (light toast)

American (heavy toast)

Oak barrel staves (16L)

French (light toast)

French (heavy toast)

What can we conclude?

A. *Brettanomyces penetrates* into oak staves.

- Approximately 8 mm.
- Affected by oak species and toasting.
- Difficult to reach with antimicrobial chemicals.

B. Steaming for 9 to 12 minutes needed to remove.

- Wineries will not know depth of penetration in any given barrel.

Research Focus
(Use of hot water)

Equivalent results to steaming if...

0 to 4 mm cross-section

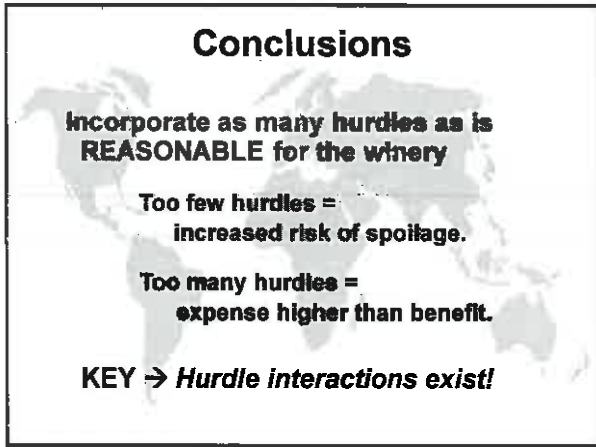
- 70°C for 28 minutes
- 80°C for 18 minutes

0 to 8 mm cross-section

- 70°C for 30 minutes
- 80°C for 20 minutes



How do we *REALLY* control *Brettanomyces*?



Conclusions

Incorporate as many hurdles as is REASONABLE for the winery

Too few hurdles = increased risk of spoilage.

Too many hurdles = expense higher than benefit.

KEY → Hurdle interactions exist!



If SO₂, filtration, heat, or chitosan does not control *Brettanomyces*...

Thank you for your support!



- **WAGrape/Wine Research Group**
- **Regional wineries**
- **Scott Laboratories**
- **Lallemand Inc.**
- **GusmerEnterprises**
- **Graduate students**

Pennsylvania Industry